Concept map

2017/18 Batch

Year 2 Semester 2.

Module Coordinator : Head Microbiology/ Head Parasitology or an appointed staff member from Department of Microbiology or Parasitology.

Resource pool

Department of Microbiology

Dr. C. Gamage (Head/Microbiology)

Prof F. Noordeen

Dr N. Dissanayake

Dr V. Liyanapathirana

Dr. C. Ratnatunga

Dr. A. Tennegedara

Department of Parasitology

Prof D. Iddawella

Dr R. Morel (Head, Parasitology)

Prof S. Wickramasingha

Dr D. Attapattu

Department of Biochemistry

Prof. Kalana Maduwage

No. of credits 4

Component	Lectures	SGDs	Practical's	Seminar	Formative assessments
					/Assignment
General	7	1	4	2	
Bacteriology	12	2	1.5		1
Virology	10	1	1.5		2
Mycology	2				
Combined Microbiology					2
Parasitology	15	3	5.5		
Total	46	12			
Credit equivalents					5

Hours of credit equivalence = 61 lecture h

Total Lectures 58h
SGD Seminars 2 h
Practical's Per student 12.5 h

Total time tabled hours 131

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Year 2 Semester 2.

Total credit hours 67.25

	Topic	Time	Objectives	Dept	Resp/person	Teaching/ Learning activity	Comments
1.	Overview of microbiology in relation to human health	1 h	Explain the interactions of microorganisms in relation to human beings State why medical undergraduates need to know about micro organisms Describe what medical microbiologists and parasitologists do	Micro	Module coordinator	Lecture	
3.	Proving causation of infection, causality - Koch's postulates and its limitations Microbial classification and visualization	1h	Discuss how causation of infections can be proved by being able to state and explain Koch's postulates and it's limitations Describe the basis of microbial classification Describe the basic structure of bacteria, fungi and viruses State the methods by which microorganisms can be visualized and identified Outline how these methods could be used to diagnose infective diseases	Micro	Module coordinator	Lecture	Includes 2 hours for facilitator meetings
2.	Introduction to Medical Parasitology and classification of parasites	1 h	List the characteristics of different groups of protozoa, helminths (nematodes, cestodes, trematodes) and arthropods	Parasit	Module coordinator	Lecture	
6.	Microscopy	1.5 h x 4 groups	Identify the parts of a compound light microscope Perform light microscopy following a standard operating	Micro and Parasit	Module coordinator	Practical	45min micro, 45min parasite

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			procedure and properly use the compound light microscope 3. Name the different types of microscopes available and their specific uses	Batch divided in to 6 groups
7. 8. 9.	Microbial growth, dissemination and survival within and outside the human host Host-parasite relationship The process by which organisms cause disease to host tissue	1 h	1. Describe the dynamics of growth in different types of micro organisms 2. List the different ways in which microorganisms survive for long periods within and outside the human host 3. Explain the terms Commensal/ normal flora, Colonizer/ transient flora and the concept of the 'microbiome' 4. Describe the relationships of microorganisms to the human host (contamination, colonization and infection) 5. List the currently known modes of transmission — microbial entry 6. Explain the terms exogenous and endogenous infections 7. Explain the term pathogenesis, 8. Describe steps that occur in establishment of microbial infections — e.g attachment, colonization, invasion, tissue damage 9. Explain how common clinical manifestations of infections reflect these mechanisms of damage in various organs	
10.	Inter-relationships among parasites, hosts and vectors	1 h	1. Define the following terms; parasite, host, endoparasite, ectoparasite, pathogen, Parasit Module Lecture coordinator	

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11. Methods of preventing infections to include sterilization and disinfection	1 h	commensal, obligatory parasite, facultative parasite, definitive host, intermediate host, reservoir host, and vectors (mechanical and biological) 2. Describe the growth, survival and spread of protozoan and helminth parasites within and outside the human host 1. Define the terms sterilization, disinfection, anti-septic and disinfectant. 2. Explain 'Standard precautions' in relation to 'Infection Control'. 3. Discuss the principles of infection prevention.	Micro	Module coordinator	Lecture	
	1 h x 8 groups	4. Perform hand hygiene, with knowledge of underlying principles, by different methods	Micro	Module coordinator	Practical	1 hour per student, 8 group per batch
Topics 1 – 11	1 h	Objectives of topics 1 - 10	Micro and Parasit	Module coordinator	SGD	
Topics 1 – 11	3h + 2 facilitator meetings	Objectives of topics 1 - 10	Micro and Parasit	Module coordinator	Seminar and 2 meetings	Includes 2 hours for facilitator meetings
Introducing medically important Bacteria						

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12. Introduction to bacteria including concepts in diagnosing bacterial infections	1h + 1.5 hr x 4 groups		 List the methods available to diagnose bacterial infections a) Visualization b) Culture and	Micro	Module coordinator	Lecture + Practical	1h + 1.5 hour per student, 4 groups per batch
13. Staphylococci	1 h	1.	Describe the medically important bacteria and their major	Micro	Module coordinator	Lecture	
14. Streptococci and Enterococci –	1h		morphological and biological characteristics that determine	Micro	Module coordinator	Lecture	
15. Gram negative cocci to include <i>Neisseria</i> and <i>Moraxella</i>	1 h	2.	visualization/ identification in the laboratory Describe the habitat, main	Micro	Module coordinator	Lecture	
16. Gram positive bacilli to include <i>Corynebacteria</i> , <i>Bacillus, Norcardia</i> and <i>Listeria</i>	1 h		mode(s) of transmission, morphology and growth characteristics in relation to identification, key virulence	Micro	Module coordinator	Lecture	
17. Mycobacteria	1 h		factors, pathogenicity and basis of clinical disease,	Micro	Module coordinator	Lecture	
18. Anaerobes including Clostridia, Actinomycetes and Prevotella	1 h	3.	Discuss the principles of laboratory diagnosis and prevention of bacterial infections	Micro	Module coordinator	Lecture	

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		4. Describe the principles of				
19. Gram negative bacilli to include enterobacteriacae , pseudomonads and other NLF of clinical importance including Acinetobacter	1 h	treatment of bacterial infections	Micro	Module coordinator	Lecture	
20. Gram negative cocco bacilli to include Haemohpilus, Bordetella, Legionella and Pasteurella	1 h		Micro	Module coordinator	Lecture	
21. Vibrio, Campylobacter and Helicobacter	1 h		Micro	Module coordinator	Lecture	
22. Spirochaetes	1 h		Micro	Module coordinator	Lecture	
23. <i>Chlamydia</i> , Rickettsioses and <i>Mycoplasma</i>	1 h		Micro	Module coordinator	Lecture	
24. Correct use of microscope, observation of smears and documentation of findings	1 h x 4 groups	 Perform light microscopy following a standard operating procedure To identify Gram positive and negative bacteria 	Micro	Module coordinator	Practical	1 hour per student, 4 groups per batch
Introducing medically important viruses						
25. Introduction to virology	1 h	Describe the general properties and classification of viruses	Micro	Module coordinator	Lecture	
26. Viruses causing Hepatitis	1 h	2. Explain the process of viral replication	Micro	Module coordinator	Lecture	
27. Pox/ adeno/ parvo/ papova viruses	1 h	3. Describe mechanisms by which viruses cause disease in humans	Micro	Module coordinator	Lecture	
28. Herpes viruses	1 h	4. State the main host defense mechanisms against viruses	Micro	Module coordinator	Lecture	
29. Respiratory viruses	1 h		Micro	Module coordinator	Lecture	

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30. Entero viruses and Viruses causing gastroenteritis	1 h		Explain the main clinical features of viral infections/disease in humans	Micro	Module coordinator	Lecture /SGD (article)	Journal article based SGD
31. Arbo viruses	1 h		Evaluate the principles of laboratory diagnosis and	Micro	Module coordinator	Lecture	
32. Retro viruses/ oncogenic viruses/ prions	1 h		prevention of viral infections Describe the principles of	Micro	Module coordinator	Lecture	
33. Viruses of zoonotic importance to include rabies	1 h	1	treatment of viral infections	Micro	Module coordinator	Lecture	
34. Diagnostics in viral infections	1 h 1.5 h x 4 groups	1.	Discuss the different methods used in the laboratory diagnosis of common viral infections a) Visualization b) Antigen detection c) Nucleic acid detection c) Culture d) Viral specific antibody detection e) Molecular detection methods to include molecular diagnosis of infectious diseases—to be included as Molecular diagnosis practical	Micro	Module coordinator	Lecture Practical	1h lecture + Practical – 1.5 h x 4 groups
35. Topics 25 to 37 MCQs	3 h		Objectives of topics 25 - 37	Micro	Module coordinator	SGD	3 SGDs, 1 hour each
Introducing medically important fungi							
36. Superficial mycoses	1h	1.	Describe fungi associated with human infections including	Micro	Module coordinator	Lecture	
37. Sub cutaneous and deep mycoses	1h		laboratory diagnosis and principals of treatment	Micro	Module coordinator	Lecture	

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38. Topics 1 - 37 – MCQs	3h	Objectives of topics 1–40	Micro	Module cordinator	SGD	3x 1 hour SGDs
39. Topics 1 – 38	2 h	Objectives of topics 1 – 40	Micro + Parasit	Module coordinator	Assignment	Training students on essay writing
		thology, pathogenesis, transmission, i				ortal of
		sitic infections in Sri Lanka and glob				
		ry diagnosis of common parasitic disc	eases in Sr	i Lanka. Explai	n the principles	s of
prevention and control of para	1				T	1
40. Malaria	1h	 Name the human malarial parasites Describe the life cycle of <i>Plasmodium</i> species Discuss the diagnostic importance of erythrocytic stages in peripheral circulation. Evaluate the methods of laboratory diagnosis Discuss the preventive measures based on the life cycle. 	Parasit	Module coordinator	Lecture	
41. Invasive intestinal protozoa - (amoebae and ciliate)	1h	Name the common intestinal amoebae and the ciliate that infect humans Outline the life cycle of Entamoeba histolytica indicating the infective, pathogenic and diagnostic stages. Describe the pathogenesis and clinical features of amoebiasis Identify points in the life cycle where preventive measures are applicable. Discuss the methods of laboratory diagnosis of the infections	Parasit	Module coordinator	Lecture	

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Year	2	Semes	ster	2.
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42. Other intestinal and urogenital protozoa – Giardia duodenalis, Cryptosporidium species and Trichomonas vaginalis	1h	1. Name the intestinal and tissue flagellates that infect humans and state their habitats 2. Name the intestinal coccidia that infect humans 3. Outline the life cycle of Giardia duodenalis, Trichomonas vaginalis and Cryptosporidum species indicating the infective,
		pathogenic and diagnostic stages. 4. Describe the pathogenesis and clinical features. 5. Evaluate the methods of laboratory diagnosis.ns 6. Discuss the preventive measures based on the life cycle
43. Haemoflagellates	1 h	1. Name the parasite(s) causing human leishmaniasis in Sri Lanka 2. Name the vector of human leishmaniasis in Sri Lanka 3. Discuss the prevention and control of leishmaniasis in Sri Lanka. 4. Describe the pathogenesis and clinical manifestations of leishmaniasis 5. Evaluate the methods of laboratory diagnosis of leishmaniasis 6. Name the parasites and vectors causing trypanosomiasis 7. Outline the geographical distribution, clinical features

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		and laboratory diagnosis of African and American trypanosomiasis				
44. Tissue coccidian (Toxoplasma gondii and Sarcocystis species)	1h	1. Name the tissue coccidia that infect humans 2. Outline the life cycle of Toxoplasma gondii indicating the infective, pathogenic and diagnostic stages. 3. Discuss the prevention and transmission of toxoplasmosis 4. Evaluate the methods of laboratory diagnosis of toxoplasmosis (congenital and acquired)	Parasit	Module coordinator	Lecture	
45. Helminths-Intestinal Nematodes Ascaris lumbricoides Necator americanus Trichuris trichiura Enterobius vermicularis Strongyloides stercoralis	2 h	Name the common intestinal nematodes in humans Describe the major morphological characteristics of parasitic nematodes Outline the life cycle of intestinal helminths indicating the infective, pathogenic and diagnostic stages. Describe the pathogenesis and clinical features. Evaluate the methods of laboratory diagnosis of intestinal nematode infections Discuss the prevention and control of intestinal nematode infections.	Parasit	Module coordinator	Lecture	

47. Cestodes and Trematodes

Taenia solium, Taenia. saginata,

Hymenolepsis diminuta,

Echinococcus granulosus

Intestinal, tissue and blood

Hymenolepsis nana,

a.Cestodes

b.Trematodes

trematodes

1 h

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46. Tissue nematodes - filarial worms	1 h	Name the major tissue nematodes of humans indicating those found in Sri Lanka. State their location in the human body.	Parasit
		 3. Name the vectors of tissue nematodes found in Sri Lanka 4. Discuss the importance of periodicity of microfilaria in relation to transmission and diagnosis. 	
		5. Outline the life cycle of Wuchereria bancrofti and Brugia malayi indicating the infective, pathogenic and diagnostic stages.	
		6. Evaluate the methods of laboratory diagnosis of filariasis7. Discuss the prevention and control filariasis	

Parasit

Module

coordinator

Lecture

1. State the major morphological

trematodes of humans

characteristics of the different

2. Outline the life cycles of cestodes

and trematodes indicating the

3. Evaluate the methods of laboratory diagnosis of cestode and trematode

4. Discuss the prevention and control.

infective, pathogenic and diagnostic stages.

infections.

groups of parasitic cestodes and

48. Arthropods of medical importance 1 - mosquitoes	1 h	Name the major mosquito borne diseases in Sri Lanka and worldwide indicating the disease(s) that they transmit Outline the life cycle of a mosquito 3. Describe the breeding habits of medically important mosquito species in Sri Lanka indicating strategies used for prevention and control.	Parasit	Module coordinator	Lecture	
49. Parasites of global importance	1h	1. Name the parasitic diseases of global importance 2. Name the-parasites causing schistosomiasis and food borne trematode infections 3. Outline the mode of transmission of schistosomes and important food-borne trematodes 4. Describe the clinical features of schistosomiasis and important food-borne trematode infections. 5. State the methods of laboratory diagnosis of these infections 6. Outline the prevention and control strategies of these infections	Parasit	Module coordinator	Lecture	
49. Arthropods of medical importance - 2	1 h	1.Define the terms mechanical and biological vectors (recall)	Parasit	Module coordinator	Lecture	
a.) Flies b)Fleas, lice and bugs c)Ticks and mites & 50. Animal bites and stings		 2. Flies Explain the importance of housefly as a mechanical vector of disease Name the other groups of flies that are medically important Describe myiasis Ticks/fleas/bugs Differentiate ticks, fleas and bugs from each other 	Parasit	Module coordinator	Lecture	

		Outline their medical importance 4. Mites Name major morphological characteristics of Sarcoptes scabiei mite Outline the life cycle 5. Lice Differentiate the three types of lice (head, body and pubic) Name the drugs used for treatment 6. Describe the prevention and control methods of vectors and ectoparasites. 1. State the common animal bites in Sri Lanka 2. State the primary and secondary effects of animal bites 3. Name the organisms that cause secondary infections of the animal bites 4. State the common marine animal and arthropod stings 5. Discuss the effects of stings				
51. Poisonous snakes and envenomation	2 h	Name the important snakes which belongs to the families-Elapidae, Viperidae and Colubridae Name the poisonous snakes in Sri Lanka Name the common (important) non-poisonous snakes in Sri Lanka	Parasit	Module coordinator	Lecture	

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		 4. Name the main phenotypic characteristics of these (2 and 3) if shown a specimen/ an image (see demonstration on snakes) 5. State the major effects of snake venom in different groups of poisonous snakes in Sri Lanka 6. State the principles underline the treatment and management of snake bites 7. State the precautions that should be taken to prevent snake bite
52. Demonstration on venomous snakes	1 hour x 4 groups	1. Identify medically important snakes of Sri Lanka if shown specimens or image Parasit Module coordinator Coordinator
53. Demonstration on intestinal protozoa and helminths	1 h x 4 groups	1. Identify pathogenic and non pathogenic intestinal amoeba, intestinal and urogenital flagellates on the stained and wet smears 2. Identify the specimens of adults and eggs of intestinal nematodes Parasit Module coordinator Coordinator Module coordinator
54. Demonstration on tissue protozoa, arthropods	1 h x 4 groups	1. Be able to identify the mosquitoes, flies fleas and mites of medical importance by their body markings 2. Identify Leishmania amastigotes on stained slide 3. Identify the trypomastigotes of trypanosomes on stained slide

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		 4. Identify malarial parasites on stained thin and thick smear 5. Identify adult taenid segments and cestode larval stages 6. Identify schistosomes adult and eggs 				
55. Practicala) Faecal smear preparation and examination for intestinal protozoan and helminthesb) Thick and thin blood smear examination for malaria parasites	1 h x 4 groups	1. Be able to properly use the compound light microscope 2. Discuss the principles regarding collection, storage and delivery/transport of faecal and blood specimens to a laboratory for diagnosis of parasitic infections 3. Be able to examine stained thin blood films and identify malaria parasites(Plasmodium falciparum and Plasmodium vivax) 4. Be able to prepare and examine wet smear of stools in saline and iodine to identify intestinal protozoal and helminth parasites	Parasit	Module coordinator	Practical	
56. Topics 40 - 55	3 h	1. Objectives of topics 40 – 55	Parasit	Module coordinator	SGD	3 x 1 hour SGDs

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Examination Format

Year 2 Semester 2.

1 h -MCQ 20 questions 3 General (Micro and Parasit)

4 Virology

5 Bacteriology

1 Mycology

7 Parasitology

2 h- SAQ (8 SAQs of 15 minutes each)

5 Microbiology 15 min questions

3 Parasitology 15 min questions

OSPE A – Parasitology 5 questions

			larks
-10	 401011	O1 11	141 170

MCQ	30%
SAQ	50%
Parasit Practical	5%
OSPE Parasit	7.5%
Micro practical	<u>7.5%</u>
	100%